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FINAL GLP REPORT: 19-02021-G1

KLIGMAN MAXIMIZATION TEST – ISO

Test Article
Latex Powder Free Gloves

21 CFR Part 58 Compliance
Good Laboratory Practice for Nonclinical Laboratory Studies

Final Report Date
8/5/2019

Study Director
Sindhura Ramasahayam, Ph.D.

Sponsor
SRI TRANG GLOVES (THAILAND) PUBLIC COMPANY LIMITED
189 Moo 7, Phlai Wat
Kanchanadit, Surat Thani 84160
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STUDY SUMMARY

The USP 0.9% Sodium Chloride for Injection (NaCl) and Cottonseed Oil (CSO) extracts of the test article, Latex Powder Free Gloves, elicited no reaction at the challenge (0% sensitization), following an induction phase. Therefore, as defined by the grading scale of the USP, the test article is classified as a non-sensitizer.

Based on the criteria of the protocol, the test article meets the requirements of the ISO 10993–10 guidelines.

QUALITY ASSURANCE STATEMENT

The Quality Assurance Unit conducted inspections on the following dates. The findings were reported to the Study Director and to Toxikon's Management.

The final report was reviewed to assure that the report accurately describes the methods and standard operating procedures. The reported results accurately reflect the raw data of the nonclinical study conducted per the protocol.

Phase	Inspection Date	Date Reported to Study Director	Date Reported to Management
DOSE ADMINISTRATION	7/3/2019	7/5/2019	7/5/2019
DATA	8/5/2019	8/5/2019	8/5/2019
FINAL REPORT	8/5/2019	8/5/2019	8/5/2019

Romel Patel, M.S.
Quality Assurance

8/5/2019
Date

GLP COMPLIANCE STATEMENT

This study meets the technical requirements of the protocol.

This study was conducted in compliance with the current U.S. Food and Drug Administration 21 CFR, Part 58 Good Laboratory Practices for Nonclinical Laboratory Studies.

The sections of the regulations not performed by or under the direction of Toxikon Corporation, exempt from this Good Laboratory Practice Statement, included characterization and stability of the test article, 21 CFR, Part 58.105, and its mixture with carriers, 21 CFR, Part 58.113.

SIGNATURES

Signature Information	
Protocol Number	p19-0828-00a
Study Director	Sindhura Ramasahayam, Ph.D.
Study Supervisor	Allan Sleger, A.S., LAT
Company	Toxikon Corporation

VERIFICATION DATES

The study initiation day is the date the protocol is signed by the Study Director.

Verification Dates	
Test Article Receipt	2/20/2019
Project Log	6/10/2019
Study Initiation	6/12/2019
Study Completion	8/5/2019

Sindhura Ramasahayam, Ph.D.
Study Director

8/5/2019
Date

1.0 PURPOSE

The purpose of the study was to determine the potential allergenic or sensitizing capacity of the test article. The study was used as a procedure for screening of contact allergens in guinea pigs and extrapolating the results to humans, but it does not establish the actual risk of sensitization.

2.0 REFERENCES

The study was based upon the following references:

- ISO 10993–10, 2010, Biological Evaluation of Medical Devices – Part 10: Tests for Irritation and Skin Sensitization.
- United States Pharmacopeia 42, National Formulary 37, 2019. <1184> Sensitization Testing.
- Zhai, H., Wilhem, K–P, and H.I. Maibach, eds. Marzulli and Maibach's Dermatotoxicology. 7th edition Boca Raton: CRC Press, 2007. 443–444, 450–451.
- Magnusson, B. and A.M. Kligman. "The Identification of Contact Allergens by Animal Assay. The Guinea Pig Maximization Test." J. Invest. Dermatol. 52 (1969): 268–276.
- Magnusson, B. and A.M. Kligman, Allergic Contact Dermatitis in the Guinea Pig. Identification of Contact Allergens. Springfield, IL.: Thomas, 1970.
- ISO 10993–12, 2012, Biological Evaluation of Medical Devices – Part 12: Sample Preparation and Reference Materials.
- ISO/IEC 17025, 2017, General Requirements for the Competence of Testing and Calibration Laboratories.

3.0 COMPLIANCE

The study conformed to the current FDA 21 CFR, Part 58 – Good Laboratory Practice for Nonclinical Laboratory Studies.

4.0 IDENTIFICATION OF TEST AND CONTROL ARTICLES

The Sponsor supplied the following information on a Test Requisition Form or other correspondence, wherever applicable (excluding confidential or trade secret information). The Sponsor was responsible for all test article characterization data as specified in the GLP regulations.

4.1 Test Article:

Name: Latex Powder Free Gloves

CAS/Code Number: Not Supplied by Sponsor (N/S)

Lot/Batch Number: 401200347

Physical State: Solid Gloves

Color: Natural-white

Expiration Date: 13/02/2022

Density: N/S

Stability: N/S

Sterility: Not Sterile

Sterilization Conditions: Non sterile

Storage Condition: Room Temperature

Safety Precautions: N/S

Intended Use: Final finished device - surface contact limited

4.2 Negative Control Articles (Toxikon Supplied):

4.2.1 Negative Control Article 1:

Name: USP 0.9% Sodium Chloride for Injection (NaCl)

Toxikon QC Number: CSC-19-05-00102; CSC-19-07-00002

4.2.2 Negative Control Article 2:

Name: Cottonseed Oil (CSO)

Toxikon QC Number: CSC-19-06-00068; CSC-19-06-00069

4.3 Positive Control Article (Toxikon Supplied):

Name: Dinitrochlorobenzene (DNCB)

Toxikon QC Number: CSC-15-09-00024

4.4 Reagents (Toxikon Supplied):

4.4.1 Reagent Name 1:

Name: Ethanol (EtOH)

Toxikon QC Number: CSC-19-03-00213

4.4.2 Reagent Name 2:

Name: USP Sterile Water for Injection (SWFI)

Toxikon QC Number: CSC-19-02-00014; CSC-18-04-00155

4.4.3 Reagent Name 3:

Name: Freund's Complete Adjuvant (FCA)

Toxikon QC Number: CSC-19-04-00036

4.4.4 Reagent Name 4:

Name: Sodium Dodecyl Sulfate (SDS)

Toxikon QC Number: CSC-18-01-00004

4.4.5 Reagent Name 5:

Name: Petrolatum

Toxikon QC Number: CSC-19-03-00039

5.0 IDENTIFICATION OF TEST SYSTEM

5.1 Animals Used in the Study:

Number and Species: 35 Hartley guinea pigs (*Cavia porcellus*)

Sex: 16 males and 19 females (females were non–pregnant and nulliparous)

Weight/Age Range: 308.9 – 400.2 grams / at least 26 days old (adult)
weighed to the nearest 0.1 g

Health Status: healthy, not previously used in other experimental procedures

Animal Purchase: Elm Hill Breeding Labs, Inc., Chelmsford, MA

Animal Identification: ear punch

Acclimation: minimum 5 days, under same conditions as for the actual test

Animal Selection: selected from larger pool and examined to ensure lack of adverse clinical signs

5.2 Animal Care and Maintenance:

Animal Room Target Temperature: 68 ± 5 °F

Animal Room Target Relative Humidity: 30–70%

Air Exchanges per Hour: a minimum of 10 changes per hour

Lights: 12–hour light/dark cycle, full spectrum fluorescent lights

Housing: group housed (per sex)

Cages: suspended stainless steel

Bedding: Alfa Cobs, ScottPharma Solutions, Marlborough, MA (non-contact)

Animal Rations: Teklad Hi–Fiber Guinea Pig Diet 2041, Envigo, Madison, WI, *ad libitum*

Water: tap water, *ad libitum*

There were no known contaminants present in the feed, water, or bedding expected to interfere with the test data.

The laboratory and animal rooms were maintained as limited–access facilities.

6.0 JUSTIFICATION OF TEST SYSTEM AND ROUTE OF ADMINISTRATION

6.1 Justification of Test System:

Historically, guinea pigs have been used in, and are generally regarded as the species of choice for, laboratory identification of skin allergens because the guidelines have no alternative (non–animal) methods.

6.2 Route of Administration:

Dermal application corresponds to a likely route of human exposure. The test article was extracted and administered *in vivo* through a medium compatible with the test system, as indicated on the Test Requisition Form.

7.0 EXPERIMENTAL DESIGN AND DOSAGE

7.1 Preparation of Test and Control Articles:

7.1.1 Preparation, Extraction Medium, and Extraction Conditions:

The test article (60.0 cm^2) was combined with 10.0 mL of vehicle following an ISO 10993–12 ratio of 6 cm^2 per 1 mL. The test article was separately extracted in NaCl and CSO at 70 ± 2 ºC for 24 ± 2 hours under dynamic conditions.

7.1.2 Addition of Extraction Medium:

Properly prepared test articles were placed in separate extraction vessels and the appropriate medium was added to each vessel. The extraction medium completely covered the test article.

7.1.3 Control Conditions:

An untreated control (blank) was prepared for parallel treatment and comparison. The untreated control was the extraction medium that was subjected to a similar incubation as used for the test article.

7.1.4 Extract Agitation:

Each extract was agitated vigorously prior to administration.

7.1.5 Extract Examination:

The test article appeared unchanged by the extraction procedure. The extracts were clear and free of particulates and the color of the vehicle unchanged.

7.1.6 Extract Manipulation:

The extracts were not filtered, centrifuged, or pH adjusted.

7.1.7 Extract Storage:

After the completion of the extraction, the extracts were kept at room temperature and were used the same day as the extraction was completed. Fresh extracts were created for each dosing phase of the study. No storage of the extracts occurred.

7.1.8 Positive Control:

The positive control, DNCB, was dissolved in 95% EtOH to a final concentration of 0.1%.

7.1.9 Other Test Article Preparation:

All other test article preparation was as specified by the Sponsor.

7.2 Pre–Dose Procedure:

The test animals were weighed and the application sites were prepared by shaving the animals with clippers to render the test sites free of any hair. On Day 0 and Day 6, a 5 cm × 7 cm area (approximate) over the shoulder region was prepared. On Day 23, a 4 cm × 4 cm area (approximate) of the flank was prepared.

7.3 Dose Administration:

7.3.1 Distribution of Animals:

(1)	Experimental	(10 animals per extract)
(2)	Negative Controls	(5 animals per extract)
(3)	Positive Controls	(5 animals per study)

7.3.2 Primary Irritation Phase:

As the test article was extracted, a Primary Irritation Phase was not performed. The extracts were used at 100% concentration for the remainder (i.e., sensitization phase) of the study.

7.3.3 Induction/Intradermal Application:

Three pairs of intradermal injections were made so that on each side of the midline there was one row of three injections each. Injections 1 and 2 were given in close proximity to each other cranially. Injection 3 was located caudally. The injection sites (6) were just within the boundaries of a 2 cm × 4 cm patch, which were applied one week following the injections. The dosing solutions were as follows:

7.3.3.1 Experimental Group (Day 0):

(1) 0.1 mL FCA 1:1 with vehicle
(2) 0.1 mL test article extract
(3) 0.1 mL test article extract 1:1 with FCA

7.3.3.2 Negative Control Group (Day 0):

(1) 0.1 mL FCA 1:1 vehicle
(2) 0.1 mL blank vehicle
(3) 0.1 mL vehicle 1:1 with FCA

7.3.3.3 Positive Control Group (Day 0):

(1) 0.1 mL FCA 1:1 NaCl
(2) 0.1 mL 0.1% DNCB in 95% EtOH
(3) 0.1 mL 0.1% DNCB in 95% EtOH 1:1 with FCA

The extracts were used neat when preparing the dosing solutions/dilutions for injection.

7.3.4 Topical Application:

On Day 6, animals that showed no signs of irritation or corrosion after the induction application were pretreated with 10% Sodium Dodecyl Sulfate (SDS) in Petrolatum 24 hours before the topical induction application. If irritation or corrosion was present, no pretreatment occurred.

7.3.4.1 Experimental Group (Day 7):

Approximately 0.3 mL of test article extract was used to "saturate" a 2 cm x 4 cm piece of absorbable material. The patch was secured with an occlusive wrapping or guinea pig jacket and left in place for 48 hours.

7.3.4.2 Negative Control Group (Day 7):

The animals were exposed to the vehicle without the test article using the same procedure utilized for the experimental group.

7.3.4.3 Positive Control Group (Day 7):

The animals were exposed to 0.1% DNCB solution in 95% EtOH, using the same procedures applied to the experimental group.

The extracts were used neat when preparing the dosing solutions/dilutions.

7.3.5 Challenge Application:

7.3.5.1 Experimental Group (Day 23):

Extract "saturated" pieces of appropriate absorbable material, measuring 2 cm × 2 cm or, a Hill Top Chamber®, was secured to a previously unexposed area of the animal for 24 hours with the same type of occlusive bandage or guinea pig jacket that was used for the Topical Induction Application. Approximately 0.3 mL of test article extract, negative control vehicle, or 0.1% DNCB in 95% EtOH was used to "saturate" the 2 cm x 2 cm piece of absorbable material or the Hill Top Chamber®.

7.3.5.2 Negative Control Group (Day 23):

For the negative control animals, the patch was saturated with the vehicles without the test article.

7.3.5.3 Positive Control Group (Day 23):

For the positive control animals, the patch was saturated with 0.1% DNCB in 95% EtOH.

The extracts were used neat when preparing the dosing solutions/dilutions.

7.4 Post Dose Procedures:

7.4.1 Skin Readings (Day 25, 26, and 27):

After removing the patches on Day 24, the challenge sites were immediately cleaned. Skin readings were taken at 24, 48, and 72 hours after the challenge exposure period (Days 25, 26, and 27). The evaluation of skin reactions used the four–point scale described in Table 1. Any animal showing a skin reaction score of 1 or greater (at any time point) was considered positive.

7.4.2 Clinical Observations:

Daily observations were made for clinical signs.

7.4.3 Scoring:

Using the Scoring System of Magnusson and Kligman (Table 1), the allergenic potential of a test article was classified based on the percent of responsive animals as described in Table 2:

TABLE 1:
Magnusson and Kligman Scale

Reaction	Grading Scale
No Visible Change	0
Discrete or Patch Erythema	1*
Moderate and Confluent Erythema	2*
Intense Erythema and Swelling	3*

* Denotes a positive response.

TABLE 2:
Sensitization Classification

Positives in Test Group (%)	Assigned Grade	Assigned Class
0	–	Nonsensitizer
< 10	1	Weak
10–30	2	Mild
31–60	3	Moderate
61–80	4	Strong
81–100	5	Extreme

The test results were interpreted based upon the percentage sensitization observed.
Note: Table 2 obtained from USP <1184>.

7.4.4 Mortality/Morbidity:

On Day 13, Animal #19 was found dead. All other animals survived the duration of the study.

7.4.5 Necropsy:

At the end of the observation period, animals were sacrificed by carbon dioxide (CO_2) inhalation.

8.0 EVALUATION CRITERIA

8.1 Evaluation of Data:

A sensitizer is a test article with which a positive response is observed in at least 10% of the test animals, as described in Table 2.

8.2 Control of Bias Statement:

The study as designed employed methodology to minimize uncertainty of measurement and to control bias for data collection and analysis, which included but was not limited to: concurrent control data, system suitability assessment, randomization, and method controls such as blanks and replicates.

9.0 RESULTS

9.1 Animal Weights (Table 3):

All animals were within the specified range of body weights (300–500 g) at the initiation of the study (Day 0).

9.2 Clinical Observations (Table 3):

Animal #19 was found dead in a cage on day 13. A gross necropsy was performed on heart, lungs, liver, spleen and kidneys and there were no abnormalities noted. The reason for death might be due to guinea pig- adenoviral infection in lungs (See Mortality report) and is unlikely related to the test article. No systemic signs of toxicity were observed in other treated or control animals.

9.3 Sensitization (Table 4):

None of the treated (NaCl or CSO extracts) or negative control animals exhibited any reaction at the challenge (0% sensitized). The positive control article elicited discrete reactions in all animals (100% sensitized).

10.0 CONCLUSION

The USP 0.9% Sodium Chloride for Injection (NaCl) and Cottonseed Oil (CSO) extracts of the test article, Latex Powder Free Gloves, elicited no reaction at the challenge (0% sensitization), following an induction phase. Therefore, as defined by the grading scale of the USP, the test article is classified as a non-sensitizer.

Based on the criteria of the protocol and these results, the test article meets the requirements of the ISO 10993–10 guidelines.

11.0 RECORDS

- Original raw data will be archived by Toxikon Corporation.
- A copy of the final report and any report amendments will be archived by Toxikon Corporation.
- The original final report and a copy of any protocol amendments or deviations will be forwarded to the Sponsor.
- The test article will be disposed by Toxikon.
- Test article retention upon study completion is the responsibility of the Sponsor.

12.0 CONFIDENTIALITY AGREEMENT

Per corporate policy, confidentiality shall be maintained in general, and in specific accordance with any relevant agreement specifically executed between Toxikon and the Sponsor.

13.0 ANIMAL WELFARE STATEMENT

The Sponsor assured that, to the best of their knowledge, this study did not unnecessarily duplicate previous testing and that there were no non–animal alternatives acceptable for the evaluation of this test article as defined by the protocol.

FCA-induced lesions at or near the FCA injection sites are an expected feature of the study protocol. FCA lesion sites judged to be large or excessive are monitored closely and reported to Veterinary Staff. The large FCA lesion sites are also recorded and reported to an Institutional Official (IO) and Institutional Animal Care and Use Committee (IACUC).

Any evidence of pain and distress was reported to the Veterinarian and/or Study Director during the course of this study.

Toxikon strictly adheres to the following standards in maintaining the animal care and use program:

United States Department of Agriculture (USDA), Animal and Plant Health Inspection Service, 9 CFR Ch. 1, Subchapter A–Animal Welfare.

“Guide for the Care and Use of Laboratory Animals,” National Research Council, 2011.

Office for Laboratory Animal Welfare (OLAW), “Public Health Service Policy on Humane Care and Use of Laboratory Animals,” Health Research Extension Act of 1985 (Public Law 99–158 November 20, 1985), revised 2015.

ISO 10993–2, 2006, Biological Evaluation of Medical Devices – Part 2: Animal Welfare Requirements.

AAALAC International accreditation.

14.0 UNFORESEEN CIRCUMSTANCES

Any unforeseen circumstances were documented in the raw data. However, no unforeseen circumstances that affected the integrity of the study were noted.

15.0 PROTOCOL AMENDMENTS/DEVIATIONS

There were no protocol amendments or deviations. No changes to the protocol were required.

TABLE 3:
Animal Weights and Clinical Observations

Group	Animal #	Sex	Body Weight (g) Day 0 7/3/2019	Signs of Toxicity*
Test Article (NaCl Extract)	1	Male	375.2	None
	2	Male	363.1	None
	3	Male	385.8	None
	4	Male	374.9	None
	5	Male	400.2	None
	6	Female	376.0	None
	7	Female	326.3	None
	8	Female	343.4	None
	9	Female	312.5	None
	10	Female	312.6	None
Test Article (CSO Extract)	11	Male	380.6	None
	12	Male	349.0	None
	13	Male	358.5	None
	14	Male	388.1	None
	15	Male	371.0	None
	16	Female	333.2	None
	17	Female	308.9	None
	18	Female	332.1	None
	19	Female	326.0	13B
	20	Female	335.7	None
Negative Control (NaCl)	21	Male	340.9	None
	22	Male	343.6	None
	23	Female	354.5	None
	24	Female	327.0	None
	25	Female	319.5	None
Positive Control (DNCB)	26	Male	340.9	None
	27	Male	378.4	None
	28	Female	339.0	None
	29	Female	333.5	None
	30	Female	361.4	None
Negative Control (CSO)	31	Male	361.4	None
	32	Male	364.3	None
	33	Female	316.5	None
	34	Female	390.7	None
	35	Female	339.5	None

* Summary of Clinical Observations - Day 0 through Day 27, excluding skin reactions.
13B: Animal found dead

TABLE 4:
Skin Examination Data

Group	Animal #	Sex	Scores Day 25 7/28/2019	Scores Day 26 7/29/2019	Scores Day 27 7/30/2019	Percent Animals Sensitized	Allergenic Potential
Test Article (NaCl Extract)	1	Male	0	0	0	0%	Non Sensitizer
	2	Male	0	0	0		
	3	Male	0	0	0		
	4	Male	0	0	0		
	5	Male	0	0	0		
	6	Female	0	0	0		
	7	Female	0	0	0		
	8	Female	0	0	0		
	9	Female	0	0	0		
	10	Female	0	0	0		
Test Article (CSO Extract)	11	Male	0	0	0	0%	Non Sensitizer
	12	Male	0	0	0		
	13	Male	0	0	0		
	14	Male	0	0	0		
	15	Male	0	0	0		
	16	Female	0	0	0		
	17	Female	0	0	0		
	18	Female	0	0	0		
	19	Female	N/A	N/A	N/A		
	20	Female	0	0	0		
Negative Control (NaCl)	21	Male	0	0	0	0%	Non Sensitizer
	22	Male	0	0	0		
	23	Female	0	0	0		
	24	Female	0	0	0		
	25	Female	0	0	0		
Positive Control (DNCB)	26	Male	1	1	1	100%	Extreme Sensitizer
	27	Male	1	1	1		
	28	Female	1	1	1		
	29	Female	1	1	1		
	30	Female	1	1	1		
Negative Control (CSO)	31	Male	0	0	0	0%	Non Sensitizer
	32	Male	0	0	0		
	33	Female	0	0	0		
	34	Female	0	0	0		
	35	Female	0	0	0		

N/A: Not Applicable

APPENDIX I:
Software Systems

Software	Use	21 CFR Part 11 Status	Publisher/ Vendor	Location
Adobe Acrobat 8, 9, and 10 Professional	Document preparation	Not Applicable	Adobe Systems, Inc.	San José, CA
Matrix Gemini 5.3.19	Laboratory Information Management System	Compliant	Autoscribe Limited	Reading, UK
MS Office 2010 Small Business Suite and MS Office 2013 Professional Suite and higher	Business software (suite includes Word, Excel, PowerPoint, Outlook, Publisher, Office tools)	Not Applicable	Microsoft Corporation	Redmond, WA
Rees Scientific Centron Presidio 3.0	Automated Environmental Monitoring	Compliant	Rees Scientific	Trenton, NJ
TMS Web 7	Document management for SOPs and training records management software system	Compliant	Quality Systems Integrators	Eagle, PA
Toxikon Protocol Manager 1.0	Protocol requisition application	Not Applicable	Toxikon Corporation	Bedford, MA

ATTACHMENT A:
MORTALITY REPORT

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CONTRIBUTING SCIENTIST REPORT

TOXIKON PROJECT NUMBER: 19-02021-G1

KLIGMAN MAXIMIZATION TEST - ISO

MORTALITY REPORT

Study Director
Sindhura Ramasahayam, Ph.D.

Contributing Scientist
Vikas Kulshreshtha, BVSc (DVM), PhD, Diplomate ACVP

Final Report Date
8/5/2019

MANAGEMENT OF THE STUDY

Performing Laboratory
Toxikon Corporation
15 Wiggins Avenue
Bedford, MA 01730

Sponsor
SRI TRANG GLOVES (THAILAND) PUBLIC COMPANY LIMITED
189 Moo 7, Phlai Wat
Kanchanadit, Surat Thani, 84160
Thailand

Animal Number(s): 19

Date of Receipt: 06/27/2019

Date of Death: 07/16/2019

1.0 HISTORY/ PHYSICAL EXAMINATION/DIAGNOSTICS:

Animal was found dead in its cage while on Kligman Maximization Test. No clinical signs were noted prior to death.

2.0 NECROPSY SUMMARY:

No gross lesions were reported to the pathologist. Heart, lungs, liver, spleen, and kidneys were collected and fixed in 10% neutral buffered formalin for routine H&E processing and histopathological analysis.

3.0 MICROSCOPIC FINDINGS:

Lungs: Multiple sections were examined. There was severe, necrotizing, bronchiolitis with desquamation of lining epithelial cells admixed with a few lymphocytes, macrophages and heterophils. The bronchiolar lumina was often occluded sloughed epithelial cells admixed with necrotic debris. The epithelial/sloughed epithelial cells often contain round to oval, basophilic viral (guinea pig adenovirus) inclusion bodies. Mild edema and fibrin with slight increase in the numbers of macrophages was observed in alveoli.

Liver: Multiple sections were examined. Multifocally, the hepatocytes exhibited clear cytoplasmic vacuolation (vacuolar degeneration, lipid type).

No significant findings were observed in other examined sections.

4.0 CONCLUSION:

The microscopic lesions in the lung were highly suggestive of guinea pig-adenoviral infection. Therefore, the cause of the death in this animal might be attributed to respiratory failure as a consequence of severe, adenoviral bronchiolitis. The hepatic vacuolar degeneration (lipid type) was considered as a spontaneous background change (E. F. McInnes 2012) but also occurs sometimes as a result of metabolic disturbances, stress or toxic insult.

REFERENCES

E. F. McInnes. Background Lesions in Laboratory Animals (A color Atlas). Saunders Elsevier, 2012.

5.0 AUTHORIZED PERSONNEL

Vikas Kulshreshtha

Vikas Kulshreshtha, BVSc (DVM), PhD, Diplomate ACVP
Pathologist

8-5-19

Date

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TOXIKON TEST PROTOCOL
FDA GLP REGULATIONS
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KLIGMAN MAXIMIZATION TEST - ISO

TOXIKON PROTOCOL NUMBER: p19-0828-00a

21 CFR Part 58 Compliance
Good Laboratory Practice for Nonclinical Laboratory Studies

MANAGEMENT OF THE STUDY

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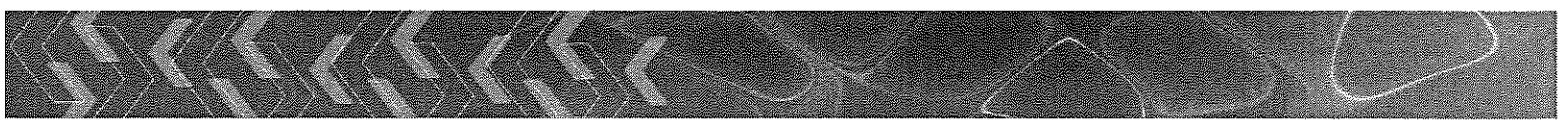

PROTOCOL SIGNATURES

TASNEE HASALEM
PRINT NAME

07/06/2019
Date

Sponsor's Representative Approval
SRI TRANG GLOVES (THAILAND) PUBLIC COMPANY LIMITED
189 Moo 7, Phlai Wat
Kanchanadit, Surat Thani 84160

Colin McFadden
PRINT NAME

6/7/19
Date

Quality Assurance Review
Toxikon Corporation
15 Wiggins Avenue
Bedford, MA 01730

Sindhura Ramasahayam
PRINT NAME

6/12/2019
Date

Study Director Signature
Toxikon Corporation
15 Wiggins Avenue
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1.0 PURPOSE

The purpose of the study is to determine the potential allergenic or sensitizing capacity of a test article. The study is used as a procedure for screening of contact allergens in guinea pigs and extrapolating the results to humans, but it does not establish the actual risk of sensitization.

2.0 REFERENCES

The study will be based upon the following references:

- ISO 10993-10, 2010, Biological Evaluation of Medical Devices - Part 10: Tests for Irritation and Skin Sensitization.
- United States Pharmacopeia 42, National Formulary 37, 2019. <1184> Sensitization Testing.
- Zhai, H., Wilhem, K-P, and H.I. Maibach, eds. Marzulli and Maibach's Dermatotoxicology. 7th edition Boca Raton: CRC Press, 2007. 443-444, 450-451.
- Magnusson, B. and A.M. Kligman. "The Identification of Contact Allergens by Animal Assay. The Guinea Pig Maximization Test." J. Invest. Dermatol. 52 (1969): 268-276.
- Magnusson, B. and A.M. Kligman, Allergic Contact Dermatitis in the Guinea Pig. Identification of Contact Allergens. Springfield, IL.: Thomas, 1970.
- ISO 10993-12, 2012, Biological Evaluation of Medical Devices - Part 12: Sample Preparation and Reference Materials.
- ISO/IEC 17025, 2017, General Requirements for the Competence of Testing and Calibration Laboratories.

3.0 COMPLIANCE

The study will conform to the current FDA 21 CFR, Part 58 - Good Laboratory Practice for Nonclinical Laboratory Studies.

4.0 IDENTIFICATION OF TEST AND CONTROL ARTICLES

The Sponsor will supply the following information on a Test Requisition Form or other correspondence, wherever applicable (excluding confidential or trade secret information). The Sponsor will be responsible for all test article characterization data as specified in the GLP regulations. Test and control articles (exclusive of extracts) that are mixed with carriers require verification of concentration, homogeneity, and stability. Samples of test and control article mixtures will be returned to the Sponsor for characterization and verification, unless this work is specifically contracted to Toxikon by Sponsor under a separate analytical protocol, whichever is applicable.

4.1 Test Article:

Name: To Be Determined (TBD)

CAS/Code Number: TBD

Lot/Batch Number: TBD

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Physical State: TBD

Color: TBD

Expiration Date: TBD

Density: TBD

Stability: TBD

Sterility: TBD

Sterilization Conditions: TBD

Storage Condition: TBD

Safety Precautions: TBD

Intended Use: TBD

4.2 Negative Control Article(s)* (Toxikon Supplied, unless specified by the Sponsor):

Name: To Be Determined (TBD)

Toxikon QC Number: TBD

* Negative control article(s) will be the vehicle(s) used for extraction, as selected by the Sponsor.

4.3 Positive Control Article(s) (Toxikon Supplied, unless specified by the Sponsor):

Name: Dinitrochlorobenzene (DNCB)

Toxikon QC Number: To Be Determined (TBD)

4.4 Reagent(s) (Toxikon Supplied, unless specified by the Sponsor):

4.4.1 Reagent Name 1:

Name: Ethanol (EtOH)

Toxikon QC Number: To Be Determined (TBD)

4.4.2 Reagent Name 2:

Name: USP Sterile Water for Injection (SWFI)

Toxikon QC Number: To Be Determined (TBD)

4.4.3 Reagent Name 3:

Name: Freund's Complete Adjuvant (FCA)

Toxikon QC Number: To Be Determined (TBD)

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4.4.4 Reagent Name 4:

Name: Sodium Dodecyl Sulfate (SDS)

Toxikon QC Number: To Be Determined (TBD)

4.4.5 Reagent Name 5:

Name: Petrolatum

Toxikon QC Number: To Be Determined (TBD)

5.0 IDENTIFICATION OF TEST SYSTEM

5.1 Animals Used in the Study:

Number and Species: 15 Hartley guinea pigs (*Cavia porcellus*), per extract, 5 Hartley guinea pigs for positive control

Sex: male and/or female (females will be non-pregnant and nulliparous)

Weight/Age Range: 300-500 grams / at least 26 days old (adult) weighed to the nearest 0.1 g

Health Status: healthy, not previously used in other experimental procedures

Animal Purchase: registered commercial breeder

Animal Identification: ear punch

Acclimation: minimum 5 days, under same conditions as for the actual test

Animal Selection: selected from larger pool and examined to ensure lack of adverse clinical signs

5.2 Animal Care and Maintenance:

Animal Room Target Temperature: 68 ± 5 °F

Animal Room Target Relative Humidity: 30-70%

Air Exchanges per Hour: a minimum of 10 changes per hour

Lights: 12-hour light/dark cycle, full spectrum fluorescent lights

Housing: group housed (per sex)

Cages: suspended stainless steel

Bedding: laboratory grade bedding used as non-contact bedding

Animal Rations: commercial guinea pig ration, *ad libitum*

Water: tap water, *ad libitum*

There will be no known contaminants present in the feed, water, or bedding expected to interfere with the test data.

The laboratory and animal rooms are maintained as limited-access facilities.

6.0 JUSTIFICATION OF TEST SYSTEM AND ROUTE OF ADMINISTRATION

6.1 Justification of Test System:

Historically, guinea pigs have been used in, and are generally regarded as the species of choice for, laboratory identification of skin allergens because the guidelines have no alternative (non-animal) methods.

6.2 Route of Administration:

Dermal application corresponds to a likely route of human exposure. The test article will be extracted and administered *in vivo* through a medium compatible with the test system, as indicated on the Test Requisition Form.

7.0 EXPERIMENTAL DESIGN AND DOSAGE

7.1 Preparation of Test and Control Articles:

7.1.1 Preparation:

The test article will be prepared at the following ratio (please indicate on the Test Requisition Form):

- According to ISO 10993-12
- No preparation required
- Sponsor–Specified

7.1.2 Extraction Medium:

The test article extracts will be prepared with the following medium (please indicate on the Test Requisition Form):

- Physiological Saline (NaCl)
- Cottonseed Oil (CSO)
- Sponsor-Specified Medium (NOTE: Extraction medium not specified by ISO 10993-12 may be required to be justified.)

7.1.3 Extraction Conditions:

The test article will be dynamically extracted (except for 121 ± 2 °C) at one of the following conditions (please indicate on the Test Requisition Form):

- 37 ± 1 °C for 72 ± 2 hours
- 50 ± 2 °C for 72 ± 2 hours
- 70 ± 2 °C for 24 ± 2 hours
- 121 ± 2 °C for 60 ± 4 minutes
- Sponsor-Specified (NOTE: Extraction conditions not specified by ISO 10993-12 may be required to be justified.)

7.1.4 Addition of Extraction Medium:

Properly prepared test article will be placed in an extraction vessel and the appropriate medium will be added, unless specified otherwise by the Sponsor. The medium should completely cover the test article, unless specified otherwise by the Sponsor.

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7.1.5 Control Conditions:

Each extraction medium (control article) will be prepared for parallel treatments and comparisons.

7.1.6 Extract Agitation:

Each extract will be agitated vigorously prior to administration.

7.1.7 Extract Examination:

Each extract will be examined for particulates and changes which may have occurred during the extraction process.

7.1.8 Extract Manipulation:

The extracts will not be pH adjusted, filtered, centrifuged, or manipulated in any way, unless requested by the Sponsor. Any post extraction manipulations will be reported and justified.

7.1.9 Extract Storage:

No storage of the extracts will occur. The extracts may be cooled to ambient conditions and will be used within 24 hours of the extraction process being completed.

7.1.10 Positive Control:

The positive control, DNCB, will be dissolved in 95% EtOH to a final concentration of 0.1%.

7.1.11 Other Test Article Preparation:

All other test article preparation will be as specified by the Sponsor.

7.2 Pre-Dose Procedure:

The test animals will be weighed and the application sites will be prepared by clipping the skin of the test site free of hair. On Day 0 and Day 6, an approximately 5 cm × 7 cm area over the shoulder region will be prepared. On Day 23, an approximately 4 cm × 4 cm area of the flank will be prepared.

7.3 Dose Administration:

7.3.1 Distribution of Animals:

(1)	Experimental	(10 animals per extract)
(2)	Negative Controls	(5 animals per extract)
(3)	Positive Controls	(5 animals per study)

7.3.2 Primary Irritation Phase:

When testing extracts, a Primary Irritation Phase will not be performed. The extracts will be used at 100% concentration.

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7.3.3 Induction/Intradermal Application:

Three pairs of intradermal injections will be made so that on each side of the midline there will be one row of three injections each. Injections 1 and 2 will be given in close proximity to each other cranially. Injection 3 will be located caudally. The injection sites (6) will be just within the boundaries of a 2 cm × 4 cm patch, which will be applied one week following the injections. The dosing solutions will be as follows:

7.3.3.1 Experimental Group (Day 0):

(1) 0.1 mL FCA 1:1 with vehicle
(2) 0.1 mL test article extract
(3) 0.1 mL test article extract 1:1 with FCA

7.3.3.2 Negative Control Group (Day 0):

(1) 0.1 mL FCA 1:1 vehicle
(2) 0.1 mL blank vehicle
(3) 0.1 mL vehicle 1:1 with FCA

7.3.3.3 Positive Control Group (Day 0):

(1) 0.1 mL FCA 1:1 NaCl
(2) 0.1 mL 0.1% DNCB in 95% EtOH
(3) 0.1 mL 0.1% DNCB in 95% EtOH 1:1 with FCA

The extract will be used neat when preparing the dosing solutions for injection.

7.3.4 Topical Application:

On Day 6, animals that show no signs of irritation or corrosion after the induction application will be pretreated with 10% Sodium Dodecyl Sulfate (SDS) in petrolatum 24 hours before the topical induction application. If irritation or corrosion is present, no pretreatment will occur.

7.3.4.1 Experimental Group (Day 7):

The test article extract will be applied to a 2 cm × 4 cm piece of an appropriate absorbable material (i.e., "patch"). Approximately 0.3 mL of test article extract will be used to "saturate" the material. The patch will be placed on the skin and secured with an occlusive wrapping or guinea pig jacket. The dressing will be left in place for 48 hours.

7.3.4.2 Negative Control Group (Day 7):

The animals will be exposed to the vehicle(s) without the test article using the same procedure as used for in the experimental group.

7.3.4.3 Positive Control Group (Day 7):

The animals will be exposed to 0.1% DNCB solution in 95% EtOH in the same manner as the experimental group.

The extract will be used neat when preparing the dosing solutions.

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7.3.5 Challenge Application:

7.3.5.1 Experimental Group (Day 23):

Extract "saturated" pieces of appropriate absorbable material, measuring 2 cm × 2 cm or a Hill Top Chamber®, will be secured to a previously unexposed area of the animal for 24 hours, with the same type of occlusive bandage or guinea pig jacket that was used for the Topical Induction Application. Approximately 0.3 mL of test article extract, negative control vehicle, or 0.1% DNCB in 95% EtOH will be used to achieve saturation.

7.3.5.2 Negative Control Group (Day 23):

For the negative control animals, the patch will be "saturated" with the vehicle(s) without the test article.

7.3.5.3 Positive Control Group (Day 23):

For the positive control animals, the patch will be "saturated" with 0.1% DNCB in 95% EtOH.

The extract will be used neat when preparing the dosing solutions/dilutions.

7.4 Post-Dose Procedures:

7.4.1 Skin Readings (Day 25, 26, and 27):

After removal of the patches on Day 24, the challenge sites will be immediately cleaned. Skin readings will be taken 24, 48, and 72 hours after the challenge exposure period (Days 25, 26, and 27, respectively). For evaluation of skin reactions a four–point scale will be used, as described in Table 1. Any animal showing a skin reaction of 1 or greater at 24, 48, or 72 hours will be considered positive.

7.4.2 Clinical Observations:

Daily observations will be made for clinical signs.

7.4.3 Scoring:

Using the scoring system of Magnusson and Kligman (Table 1), the allergenic potential of a test article may be classified based on the percent of responsive animals as described in Table 2:

TABLE 1:
Magnusson and Kligman Scale

Reaction	Grading Scale
No Visible Change	0
Discrete or Patch Erythema	1*
Moderate and Confluent Erythema	2*
Intense Erythema and Swelling	3*

* Denotes a positive response.

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TABLE 2:
Sensitization Classification

Positives in Test Group (%)	Assigned Grade	Assigned Class
0	–	Nonsensitizer
< 10	1	Weak
10-30	2	Mild
31-60	3	Moderate
61-80	4	Strong
81-100	5	Extreme

The test results will be interpreted based upon the percentage sensitization observed.
Note: Table 2 adapted from USP <1184>.

7.4.4 Mortality/Morbidity:

In the event that an animal is found dead or moribund during the course of the study, the animal will be humanely sacrificed if necessary, and a gross necropsy will be performed at the earliest convenience (any abnormal tissues will be collected for histopathological analysis). If it can be identified that the morbidity or death was unrelated to the test article, the animal may be replaced after consulting the Sponsor. If a single animal, within a dose group, expires during the study, it will not be replaced. If two or more animals expire, they will be replaced with a sufficient number of animals as to ensure that the appropriate group sizes complete the study.

7.4.5 Necropsy:

At the end of the observation period, animals will be sacrificed by carbon dioxide (CO_2) inhalation.

8.0 EVALUATION CRITERIA

8.1 Evaluation of Data:

A sensitizer is a test article with which a positive response is observed in at least 10% of the test animals, as described in Table 2.

8.2 Control of Bias Statement:

The study as designed employs methodology to minimize uncertainty of measurement and to control bias for data collection and analysis, which includes but is not limited to: control data (retrospective, concurrent, or prospective), system suitability assessment, randomization, method controls such as blanks and replicates, or others as required by the specific study or guideline. Methods employed will be specified in the final report.

9.0 RECORDS

- Original raw data will be archived by Toxikon Corporation.
- A copy of the final report and any report amendments will be archived by Toxikon Corporation.
- The original final report and a copy of any protocol amendments or deviations will be forwarded to the Sponsor.

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- All used and unused test article will be handled as specified on the Test Requisition Form. If not indicated on the Test Requisition Form, all remaining test article will be disposed.
- Test article retention upon study completion is the responsibility of the Sponsor.

10.0 CONFIDENTIALITY AGREEMENT

Per corporate policy, confidentiality will be maintained in general, and in specific accordance with any relevant agreement specifically executed between Toxikon and the Sponsor.

11.0 ANIMAL WELFARE STATEMENT

The Sponsor assures that, to the best of their knowledge, this study does not unnecessarily duplicate previous testing and that there are no non-animal alternatives acceptable for the evaluation of the test article as defined by the protocol.

Evidence of pain and distress will be immediately reported to the Veterinarian and/or Study Director, who will make a decision, independently or in concert with the Sponsor, to terminate the study or to continue with or without appropriate analgesics. In toxicity studies, animals cannot be administered analgesics since they would interfere with the toxicity determination. Animals may be immediately euthanized. In other studies, one or more analgesics may be administered to reduce pain and distress. The Institutional Official (IO) and the Institutional Animal Care and Use Committee (IACUC) bases this policy upon Toxikon's Standard Operating Procedures and animal care and welfare standards as governed.

Toxikon strictly adheres to the following standards, where applicable, in maintaining the animal care and use program:

United States Department of Agriculture (USDA), Animal and Plant Health Inspection Service, 9 CFR Ch. 1, Subchapter A-Animal Welfare.

"Guide for the Care and Use of Laboratory Animals," National Research Council, 2011.

Office for Laboratory Animal Welfare (OLAW), "Public Health Service Policy on Humane Care and Use of Laboratory Animals," Health Research Extension Act of 1985 (Public Law 99-158 November 20, 1985), revised 2015.

ISO 10993-2, 2006, Biological Evaluation of Medical Devices - Part 2: Animal Welfare Requirements.

AAALAC International accreditation.

12.0 UNFORESEEN CIRCUMSTANCES

All unforeseen circumstances will be documented in the raw data. Any unforeseen circumstances that affect the integrity of the study will be discussed in the final report.

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13.0 PROTOCOL AMENDMENTS/DEVIATIONS

All changes to the approved protocol and the reason for the changes will be documented in writing, signed by the Study Director, dated, and maintained with the protocol. A Protocol Amendment (PA) or a Protocol Deviation (PD) will be generated as closely as possible to the time of the change. The document will be created and signed by the Study Director and sent to the Sponsor. Sponsor's signature will be required for amendments (PA) to indicate approval of the amendment. Acknowledgement of notification of deviations is preferred and may be with a signature or other form of documentation.

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APPENDIX I:
Software Systems

The following are the proposed software systems to be used during the conduct of this study. The actual systems used, as well as 21 CFR Part 11 compliance if applicable, will be documented in the final report

Software	Use	Publisher/ Vendor	Location
Adobe Acrobat 8, 9, and 10 Professional	Document preparation	Adobe Systems, Inc.	San José, CA
Matrix Gemini 5.3.19	Laboratory Information Management System	Autoscribe Limited	Reading, UK
MS Office 2010 Small Business Suite and MS Office 2013 Professional Suite and higher	Business software (suite includes Word, Excel, PowerPoint, Outlook, Publisher, Office tools)	Microsoft Corporation	Redmond, WA
Rees Scientific Centron Presidio 3.0	Automated Environmental Monitoring	Rees Scientific	Trenton, NJ
Report Automation 1.0	Custom software (add-in) for final report generation, review, approval, distribution to sponsors, and storage	Court Square Group	Springfield, MA
TMS Web 7	Document management for SOPs and training records management software system	Quality Systems Integrators	Eagle, PA
Toxikon Protocol Manager 1.0	Protocol requisition application	Toxikon Corporation	Bedford, MA

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